

| | | |
|--|---|---|
| FORM PTO-1390 U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371 | | ATTORNEY'S DOCKET NUMBER: 53631-65307 <div style="border: 1px solid black; padding: 2px; display: inline-block; margin-top: 5px;"> 09/914352 </div> |
| INTERNATIONAL APPLICATION NO.: PCT/SE00/00346 | INTERNATIONAL FILING DATE: 22 FEBRUARY 2000 | PRIORITY DATE CLAIMED: 25 FEBRUARY 1999 |
| TITLE OF INVENTION: MITE PROTEIN | | |
| APPLICANT(S) FOR DO/EO/US: Jens MATSSON | | |
| Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information: | | |
| 1. <input checked="" type="checkbox"/> | This is a FIRST submission of items concerning a filing under 35 U.S.C. 371. | |
| 2. <input type="checkbox"/> | This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371. | |
| 3. <input checked="" type="checkbox"/> | This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1). | |
| 4. <input checked="" type="checkbox"/> | A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. | |
| 5. <input checked="" type="checkbox"/> | A copy of the International Application as filed (35 U.S.C. 371(c)(2)) | |
| a. <input checked="" type="checkbox"/> | is transmitted herewith (required only if not transmitted by the International Bureau). | |
| b. <input type="checkbox"/> | has been transmitted by the International Bureau. (see attached copy of PCT/IB/308) | |
| c. <input type="checkbox"/> | is not required, as the application was filed in the United States Receiving Office (RO/US). | |
| 6. <input type="checkbox"/> | A translation of the International Application into English (35 U.S.C. 371(c)(2)). | |
| 7. <input type="checkbox"/> | Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)). | |
| a. <input type="checkbox"/> | are transmitted herewith (required only if not transmitted by the International Bureau). | |
| b. <input type="checkbox"/> | have been transmitted by the International Bureau. | |
| c. <input type="checkbox"/> | have not been made; however, the time limit for making such amendments has NOT expired. | |
| d. <input type="checkbox"/> | have not been made and will not be made. | |
| 8. <input type="checkbox"/> | A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)). | |
| 9. <input type="checkbox"/> | An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). | |
| 10. <input type="checkbox"/> | A translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)). | |
| Item 11. to 16. below concern document(s) or information included: | | |
| 11. <input type="checkbox"/> | An Information Disclosure Statement under 37 CFR 1.97 and 1.98. | |
| 12. <input type="checkbox"/> | An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included. | |
| 13. <input checked="" type="checkbox"/> | A FIRST preliminary amendment. | |
| 14. <input type="checkbox"/> | A SECOND or SUBSEQUENT preliminary amendment. | |
| 15. <input type="checkbox"/> | A substitute specification. | |
| 16. <input type="checkbox"/> | A change of power of attorney and/or address letter. | |
| 16. <input checked="" type="checkbox"/> | Other items or information: | |
| INTERNATIONAL PRELIMINARY EXAMINATION REPORT (PCT/IPEA/409), INTERNATIONAL SEARCH REPORT (PCT/ISA/210), ABSTRACT on a separate sheet, SEQUENCE LISTING, APPLICATION DATA SHEET | | |

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)

09/914352

INTERNATIONAL APPLICATION NO.
PCT/SE00/00346ATTORNEY'S DOCKET NO.
53631-6530717. ☒ The following fees are submitted:**BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):**

Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$ 1,000.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$ 860.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 710.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$ 690.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$ 100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =

CALCULATIONS PTO USE ONLY

Surcharge of \$130.00 for furnishing the oath or declaration later than 30 months from the earliest claimed priority date (37 CFR 1.492(e)).

| CLAIMS | NUMBER FILED | NUMBER EXTRA | RATE | \$ |
|--|--------------|--------------|------------|----------|
| Total claims | 24 - 20 = | 4 | X \$18.00 | \$ 72.00 |
| Independent claims | 1 - 3 = | 0 | X \$80.00 | \$ |
| MULTIPLE DEPENDENT CLAIMS(S) (if applicable) | | | + \$270.00 | \$ |

TOTAL OF ABOVE CALCULATIONS =

Reduction of ½, if applicant is entitled to Small Entity status under 37 CFR 1.27. +

SUBTOTAL =

Processing fee of \$130 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.49(f)).

TOTAL NATIONAL FEE =

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +

TOTAL FEES ENCLOSED =Amount to be
refunded:

charged:

- a. ☒ A check in the amount of \$ 1,202.00 to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. **25-0120** in the amount of \$ to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required by 37 CFR 1.16 and 1.17, or credit any overpayment to Deposit Account No. **25-0120**. A duplicate copy of this sheet is enclosed.

SEND ALL CORRESPONDENCE TO:

CUSTOMER No. 00466
 YOUNG & THOMPSON
 745 South 23rd Street
 2nd Floor
 Arlington, VA 22202
 (703) 521-2297
 facsimile (703) 685-0573

August 27, 2001

By

Benoit Castel
 Benoit Castel
 Attorney for Applicant
 Registration No. 35,041

Rec'd PCT/PTO 14 DEC 2001

PATENTS

#5/B

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

Jens MATTSSON

Serial No. 09/914,352

BOX PCT

Filed February 22, 2000

MITE PROTEIN

PRELIMINARY AMENDMENT

Commissioner for Patents

Washington, D.C. 20231

Sir:

Prior to issuance of an Official Action on the merits, please amend the above-identified application as follows:

IN THE SPECIFICATION:

Replace the paragraph beginning at page 11, line 17, with the following rewritten paragraph:

--In an especially advantageous embodiment, the protein according to the invention is a recombinant protein, which in addition to a functional part or all of the sequence disclosed in SEQ ID NO. 2 also comprises a tag, such as a conventional further amino acid sequence, which confers properties that facilitates purification, downstream analysis, such as Western blot, reversible immobilization, immunoprecipitation, immunofluorescence analysis etc. Said tag may e.g. be the peptide His6(Piece of SEQ ID NO: 3). In a particular embodiment, the present invention is a fusion protein, wherein the present

1044660

protein or a functional subsequence thereof is fused with another protein, such as β -galactosidase, glutathione-S-transferase, protein A etc. In the context of fusion proteins, see e.g. Smith and Johnson (1988) *Gene* 67:31; Hopp et al. (1988) *Biotechnology* 6:1204; La Vallie et al. (1993) *Biotechnology* 11:187.--

Replace the paragraph beginning at page 22, line 1, with the following rewritten paragraph:

--Competitive assay formats are preferred in the present context, wherein the amount of analyte, preferably an unknown quantity of antibodies in a subject, in a sample is measured indirectly by measuring the amount of added analyte, displaced from a capture agent by the analyte present in the sample. Most preferred are the enzyme-linked immunosorbent assay (ELISA) methods, in which an antibody typically is bound to an enzyme, such as peroxidase or phosphatase, which can produce colored reaction products from an appropriate buffer. Thus, it utilizes a tagged antigen molecule of known quantity to determine an unlabelled antigen of unknown quantity. Preferably, the protein according to the invention, or a suitable functional fragment thereof, is used coupled to a conventional tag, such as His6(Piece of SEQ ID NO: 3). This assay is e.g. useful to diagnose *Sarcoptes scabiei* infection in dogs.--

Replace the paragraph beginning at page 27, line 4, with the following rewritten paragraph:

--Cloning of part of the 5' cDNA end of MSA1

A PCR strategy was used in order to clone regions upstream of the cDNA insert in pPU3. In the first PCR the primer KBE 5 (5'CAC TAT CGG AGA ACG TAA CTT CGG 3') (SEQ ID NO: 4), complementary to the anti-sense strand of the insert in pPU3, was designed and used together with a T3 primer, complementary to the vector used to construct the cDNA library. As a template the *S. scabiei* cDNA-library was used. The resulting fragment was cloned into the *Sma*I-site of pUC18 and sequenced as above. This new fragment was then used for the design of a second primer KBE 8 (5'CCT GGC ATT CTA CTT GAG ATG TA 3') (SEQ ID NO: 5) for the amplification an additional 5'end cDNA fragment. The second 5'end fragment was cloned and sequenced as above. A continuous cDNA which included the original MSA1 cDNA and both of the 5' end fragments was generated by using the Titan™ One Tube RT-PCR system (manufactured by Roche). For the reverse transcriptase step the reverse primer MSA1Xba (5'CGC **TCT AGA** CTC AAC AAT GAA TGT CTG CAA 3') (SEQ ID NO: 6) was used. In the PCR, the reverse primer was used in combination with the forward primer LDL 2 (5'CGG **GAT CCG** AAT ATT TCG TCT CGA AAC CG 3') (SEQ ID NO: 7). The resulting fragment was cloned into the *Bam*HI-*Xba*I sites of pPU16 utilizing the recognition sites introduced during the PCR (shown in boldface). A graphic overview of the cloning strategy is shown in Fig.--

MATTSSON S.N. 09/914,352

Attached hereto is a marked-up version of the changes made to the specification. The attached page is captioned "VERSION WITH MARKINGS TO SHOW CHANGES MADE."

Respectfully submitted,

YOUNG & THOMPSON

By



Robert J. Patch
Attorney for Applicant
Registration No. 17,355
745 South 23rd Street
Arlington, VA 22202
Telephone: 521-2297

December 14, 2001

RECEIVED

VERSION WITH MARKINGS TO SHOW CHANGES MADE

IN THE SPECIFICATION:

Paragraph beginning at line 17 of page 11 has been amended as follows:

In an especially advantageous embodiment, the protein according to the invention is a recombinant protein, which in addition to a functional part or all of the sequence disclosed in SEQ ID NO. 2 also comprises a tag, such as a conventional further amino acid sequence, which confers properties that facilitates purification, downstream analysis, such as Western blot, reversible immobilization, immunoprecipitation, immunofluorescence analysis etc. Said tag may e.g. be the peptide His6(Piece of SEQ ID NO: 3). In a particular embodiment, the present invention is a fusion protein, wherein the present protein or a functional subsequence thereof is fused with another protein, such as β -galactosidase, glutathione-S-transferase, protein A etc. In the context of fusion proteins, see e.g. Smith and Johnson (1988) *Gene* 67:31; Hopp et al. (1988) *Biotechnology* 6:1204; La Vallie et al. (1993) *Biotechnology* 11:187.

Paragraph beginning at line 1 of page 22 has been amended as follows:

Competitive assay formats are preferred in the present context, wherein the amount of analyte, preferably an unknown

quantity of antibodies in a subject, in a sample is measured indirectly by measuring the amount of added analyte, displaced from a capture agent by the analyte present in the sample. Most preferred are the enzyme-linked immunosorbent assay (ELISA) methods, in which an antibody typically is bound to an enzyme, such as peroxidase or phosphatase, which can produce colored reaction products from an appropriate buffer. Thus, it utilizes a tagged antigen molecule of known quantity to determine an unlabelled antigen of unknown quantity. Preferably, the protein according to the invention, or a suitable functional fragment thereof, is used coupled to a conventional tag, such as His6(Piece of SEQ ID NO: 3). This assay is e.g. useful to diagnose *Sarcoptes scabiei* infection in dogs.

Paragraph beginning at line 4 of page 27 has been amended as follows:

Cloning of part of the 5' cDNA end of MSA1

A PCR strategy was used in order to clone regions upstream of the cDNA insert in pPU3. In the first PCR the primer KBE 5 (5'CAC TAT CGG AGA ACG TAA CTT CGG 3') (SEQ ID NO: 4), complementary to the anti-sense strand of the insert in pPU3, was designed and used together with a T3 primer, complementary to the vector used to construct the cDNA library. As a template the *S. scabiei* cDNA-library was used. The resulting fragment was cloned into the *Sma*I-site of pUC18 and sequenced as above. This new fragment was then

used for the design of a second primer KBE 8 (5'CCT GGC ATT CTA CTT GAG ATG TA 3') (SEQ ID NO: 5) for the amplification an additional 5'end cDNA fragment. The second 5'end fragment was cloned and sequenced as above. A continuous cDNA which included the original MSA1 cDNA and both of the 5' end fragments was generated by using the Titan™ One Tube RT-PCR system (manufactured by Roche). For the reverse transcriptase step the reverse primer MSA1Xba (5'CGC **TCT AGA** CTC AAC AAT GAA TGT CTG CAA 3') (SEQ ID NO: 6) was used. In the PCR, the reverse primer was used in combination with the forward primer LDL 2 (5'**CGG GAT CCG** AAT ATT TCG TCT CGA AAC CG 3') (SEQ ID NO: 7). The resulting fragment was cloned into the *Bam*HI-*Xba*I sites of pPU16 utilizing the recognition sites introduced during the PCR (shown in boldface). A graphic overview of the cloning strategy is shown in Fig.

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

Jens MATSSON

Box PCT

Serial No. (unknown)
(PCT/SE00/00346)

Application Branch

Filed herewith

MITE PROTEIN

#3/A

PRELIMINARY AMENDMENT

Commissioner for Patents

Washington, D.C. 20231

Sir:

Prior to the first Official Action and calculation of the filing fee, please amend the above-identified application as follows:

IN THE CLAIMS:

Amend claim 4 as follows:

--4. (Amended) An isolated nucleic acid encoding a protein according to claim 1.--

Amend claim 6 as follows:

--6. (Amended) A nucleic acid which hybridizes specifically under stringent conditions to a nucleic acid according to claim 4.--

Amend claim 7 as follows:

--7. (Amended) An expression vector which comprises a nucleic acid according to claim 4.--

Amend claim 8 as follows:

--8. (Amended) A method for producing a protein, which method comprises the steps of

09/914352

- (a) providing a DNA according to claim 4;
- (b) introducing said DNA in an expression vector;
- (c) insertion of said vector into a suitable host cell;
- (d) culturing said host cell to obtain the desired protein product; and optionally
- (e) purification of the protein or polypeptide produced.--

Amend claim 10 as follows:

--10. (Amended) An antibody raised against a protein according to claim 1.--

Amend claim 12 as follows:

--12. (Amended) Use of a protein according to claim 1 in an immunosorbent assay, such as enzyme-linked immunosorbent assay (ELISA).--

Amend claim 13 as follows:

--13. (Amended) Use of a protein according to claim 1 in a screening method wherein compounds having the same or similar biological activities as said protein are identified.--

Amend claim 14 as follows:

--14. (Amended) a method for screening protein or peptide analogues that mimic at least a part of the structure of the protein according to claim 1, which comprises the steps of

- (a) producing a multiplicity of analogue structures and
- (b) selecting an analogue structure, wherein the three-dimensional configuration and spatial arrangement of one or

more biologically active regions remain substantially preserved.--

Amend claim 16 as follows:

--16. (Amended) A protein according to claim 1 for use as a vaccine.--

Amend claim 17 as follows:

--17. (Amended) Use of a protein according to claim 1 in the manufacture of a vaccine preparation.--

Amend claim 18 as follows:

--18. (Amended) A vaccine preparation comprising a protein according to claim 1 and a pharmaceutically and/or veterinary acceptable carrier.--

Amend claim 20 as follows:

--20. (Amended) A method of preventing a disease associated with mites, such as *Sarcoptes scabiei*, in a subject, such as a human, canine or porcine subject, which method comprises administration of a preparation according to claim 18 to said subject in a pharmaceutically effective dose.--

Amend claim 22 as follows:

--22. (Amended) A method for the diagnosis of a mite associated disease comprising the steps of
(a) immobilizing a protein according to claim 1;
(b) providing a sample suspected of being infected with said mite associated disease;
(c) incubation of said sample with said immobilized protein;
and

(d) detection of an antibody bound to the immobilized antigen and thus specific for said mite associated disease; whereby a conclusion regarding the diagnosed condition is obtained.--

Amend claim 24 as follows:

--24. (Amended) A kit for performing the method according to claim 23.--

R E M A R K S

The above changes in the claims merely place this national stage application in the same condition as it was during Chapter II of the international stage, with the multiple dependencies being removed.

Attached hereto is a marked-up version of the changes made to the claims by the current amendment. The attached page is captioned "VERSION WITH MARKINGS TO SHOW CHANGES MADE."

Respectfully submitted,

YOUNG & THOMPSON

By Benoît Castel
Benoît Castel
Attorney for Applicant
Registration No. 35,041
Customer No. 00466
745 South 23rd Street
Arlington, VA 22202
Telephone: 703/521-2297

August 27, 2001

VERSION WITH MARKINGS TO SHOW CHANGES MADE

Amend claim 4 as follows:

--4. (Amended) An isolated nucleic acid encoding a protein according to ~~any one of claims 1-3~~ claim 1.--

Amend claim 6 as follows:

--6. (Amended) A nucleic acid which hybridizes specifically under stringent conditions to a nucleic acid according to claim 4 ~~or 5~~.--

Amend claim 7 as follows:

--7. (Amended) An expression vector which comprises a nucleic acid according to ~~any one of claims 4-6~~ claim 4.--

Amend claim 8 as follows:

--8. (Amended) A method for producing a protein, which method comprises the steps of

- (a) providing a DNA according to ~~any one of claims 4-6~~ claim 4;
- (b) introducing said DNA in an expression vector;
- (c) insertion of said vector into a suitable host cell;
- (d) culturing said host cell to obtain the desired protein product; and optionally
- (e) purification of the protein or polypeptide produced.--

Amend claim 10 as follows:

--10. (Amended) An antibody raised against a protein according to ~~any one of claims 1-3~~ claim 1.--

Amend claim 12 as follows:

--12. (Amended) Use of a protein according to ~~any one of claims 1-3~~ claim 1 in an immunosorbent assay, such as enzyme-linked immunosorbent assay (ELISA).--

Amend claim 13 as follows:

--13. (Amended) Use of a protein according to ~~any one of claims 1-3~~ claim 1 in a screening method wherein compounds having the same or similar biological activities as said protein are identified.--

Amend claim 14 as follows:

--14. (Amended) a method for screening protein or peptide analogues that mimic at least a part of the structure of the protein according to ~~any one of claims 1-3~~ claim 1, which comprises the steps of

- (a) producing a multiplicity of analogue structures and
- (b) selecting an analogue structure, wherein the three-dimensional configuration and spatial arrangement of one or more biologically active regions remain substantially preserved.--

Amend claim 16 as follows:

--16. (Amended) A protein according to ~~any one of claims 1-3~~ claim 1 for use as a vaccine.--

Amend claim 17 as follows:

--17. (Amended) Use of a protein according to ~~any one of claims 1-3~~ claim 1 in the manufacture of a vaccine preparation.--

Amend claim 18 as follows:

--18. (Amended) A vaccine preparation comprising a protein according to ~~any one of claims 1-3~~ claim 1 and a pharmaceutically and/or veterinary acceptable carrier.--

Amend claim 20 as follows:

--20. (Amended) A method of preventing a disease associated with mites, such as *Sarcoptes scabiei*, in a subject, such as a human, canine or porcine subject, which method comprises administration of a preparation according to claim 18 ~~or 19~~ to said subject in a pharmaceutically effective dose.--

Amend claim 22 as follows:

--22. (Amended) A method for the diagnosis of a mite associated disease comprising the steps of

(a) immobilizing a protein according to ~~any one of claims 1-3~~ claim 1;

(b) providing a sample suspected of being infected with said mite associated disease;

(c) incubation of said sample with said immobilized protein; and

(d) detection of an antibody bound to the immobilized antigen and thus specific for said mite associated disease; whereby a conclusion regarding the diagnosed condition is obtained.--

Amend claim 24 as follows:

--24. (Amended) A kit for performing the method according to claim ~~22 or 23~~ --

ABSTRACT OF THE DISCLOSURE

A novel major mite antigen has been isolated and sequenced for the first time. The antigen is a protein originating from the mite *Sarcoptes scabiei*. An encoding nucleic acid and advantageous uses of the novel protein and/or functional fragments thereof, e.g. in immunological testing, such as in ELISA methods, as well as in the preparation of vaccines are also disclosed.

MITE PROTEIN

Technical field

The present invention relates to a novel recombinant protein useful in the diagnosis of mite infections in mammals, such as dogs. The invention also relates to a nucleic acid encoding the protein according to the invention as well as to various advantageous uses thereof.

Background

Scabies or sarcoptic mange, which is caused by infection with the parasitic mite *Sarcoptes scabiei*, is a widespread, highly contagious disease. The parasite has been found in well over 40 different mammals, including man. During the infection, female mites burrow in the skin to feed and their activities cause an intense irritation that leads to itching and scratching, which may aggravate the condition. Sensitization of the host to the mites and their products probably plays an important role in the pathogenesis of the disease. In its extreme form, scabies can develop into a severe hyperkeratotic form, where several thousands of mites can be found in the lesions. This form of crusted scabbies is also frequently observed among immune compromised individuals, i.e. HIV patients.

The standard method for diagnosing scabies include microscopic detection of the mites and their eggs and feces in skin scrapings. This method is relatively time-consuming and, in many cases, the results of microscopic examination can be negative owing to the low number of parasites present in each sample. The use of enzyme-linked immunosorbent assays (ELISA) for the detection of antibodies to *S. scabiei* has been reported for several different animals including humans (Bornstein & Zakrisson (1993) *Vet. Dermatol.* 4:107; Arlian *et al.* (1994) *Exp. Parasitol.* 78:37; Normaznah *et al.* (1996) *Southeast Asian J. Trop. Med. Public Health* 27:53; and Hollanders *et al.* (1997) *Vet. Parasitol.* 69:117). The ELISA technology offers several distinct advantages compared to skin scrapings: A relative ease of sampling, the potential to screen large numbers of samples and the possibility to standardize

reagents and methods. The only major drawback is the limited amount of antigenic material available due to the lack of an *in vitro* propagation system for *S. scabiei*.

Arlian and colleagues (J. Med. Entomol. (1988) 25:52) have developed an *in vivo* propagation system after establishing *S. scabiei* var. *canis* on rabbits. However, the parasite burden is still relatively low and the method cannot be applied for the production of antigens for large scale screening projects. Mites from naturally infected red foxes have been successfully isolated in large numbers and used in ELISAs both for dogs and pigs. The difficulties involved in using wild foxes as a source for antigen supply are however evident.

This lack of material has not only limited large scale screening efforts and control programs among farm animals, but it has also limited the possibilities to study other aspects of scabies, for instance pathogenesis.

EP 0 473 111 provides a recombinant mite allergen which is effective as a therapeutic agent and diagnostic reagent for mite allergic diseases. However, the mite allergen of this reference are derived from a house dust mite, more specifically from *Dermatophagoides farinae*. Thus, firstly, the said house dust mite is found free-living in a house-hold environment and does not live as a parasite on a host. Thus, disease thereof is not the result of an infection but the inhalation of mites or mite body fragments or faeces. The inhalation of house dust mites may cause allergic reactions in susceptible individuals resulting in respiratory distress, such as asthma and rhinitis. Accordingly, EP 0 473 111 relates to molecules useful in the diagnosis and prevention of such conditions.

Summary of the invention

According to the present invention, the problems defined above are solved by providing a novel major mite antigen, which has now been isolated and sequenced for the first time. Thus, the invention relates an isolated antigenic protein as well as

to the nucleic acid said novel protein, as defined in the appended claims. Further, the invention also relates to various advantageous uses of the novel protein and functional fragments thereof, e.g. in immunological testing, such as in ELISA methods.

Brief description of the drawings

Figure 1 shows the extent of the recombinant protein according to the invention in relation to the native protein.

Figure 2 illustrates the recombinant plasmid pPU17 used for the expression of the mite protein according to the invention.

Figure 3 is an overview of the cloning strategy of the 5' end of MSA.

Figure 4 shows the results of Western blot analysis of the mite recombinant protein according to the invention.

Definitions

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers.

The term "nucleic acid" refers to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogs of natural nucleotides that can function in a similar manner as naturally occurring nucleotides.

The term "an analogue or functional fragment" refers to a compound which exhibits antigenic properties essentially equivalent to the ones of the protein according to the invention.

5 The phrase "hybridizing specifically to", refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (*e.g.*, total cellular DNA or RNA. The term "stringent conditions" refers to conditions under which a probe will hybridize to its target subsequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point T_m for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH, and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. (As the target sequences are generally present in excess, at T_m , 50% of the probes are occupied at equilibrium). Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide.

20 A "label" is a composition detectable by spectroscopic, photochemical, biochemical, immunochemical, or chemical means. For example, useful labels include ^{32}P , fluorescent dyes, electron-dense reagents, enzymes (*e.g.*, as commonly used in a ELISA), biotin, dioxigenin, or haptens and proteins for which antisera or monoclonal antibodies are available.

25 As used herein a "nucleic acid probe" is defined as a nucleic acid capable of binding to a target nucleic acid of complementary sequence through one or more types of chemical bonds, usually through complementary base pairing, usually through hydrogen bond formation. As used herein, a probe may include natural (*i.e.* A, G, C, U or T) or modified bases (7-deazaguanosine, inosine, *etc.*) In addition, the bases in

30

a probe may be joined by a linkage other than a phosphodiester bond, so long as it does not interfere with hybridization.

5 A "labeled nucleic acid probe" is a nucleic acid probe that is bound, either covalently, through a linker, or through ionic, van der Waals or hydrogen bonds to a label such that the presence of the probe may be detected by detecting the presence of the label bound to the probe.

10 The term "target nucleic acid" refers to a nucleic acid (often derived from a biological sample), to which a nucleic acid probe is designed to specifically hybridize. It is either the presence or absence of the target nucleic acid that is to be detected, or the amount of the target nucleic acid that is to be quantified. The target nucleic acid has a sequence that is complementary to the nucleic acid sequence of the corresponding probe directed to the target. The term target nucleic acid may
15 refer to the specific subsequence of a larger nucleic acid to which the probe is directed or to the overall sequence (e.g., gene or mRNA) whose expression level it is desired to detect. The difference in usage will be apparent from context.

20 "Subsequence" refers to a sequence of nucleic acids or amino acids that comprise a part of a longer sequence of nucleic acids or amino acids (e.g., polypeptide), respectively.

25 "Combinatorial library" means a library of molecules containing a large number, typically between 10^3 and 10^6 , of different-sequence oligomers, typically characterized in having different sequences or subunits, or a combination of different sequences of side chains and linkages, or different-substituent compounds in a small-compound library.

30 "Different-sequence oligomer compounds" are oligomers, such as oligonucleotides, oligonucleotide analogs, oligopeptides, oligopeptide analogs, oligosaccharides, or lipopeptides with different permutations of lipid and/or sequences in the peptide

moieties, glycopeptides with different sequence permutations in the saccharide and/or peptide moieties, non-biological oligomers with different-sequence permutations, or different-substituent compounds in a small-compound library.

5 The term "recombinant" when used with reference to a cell, or nucleic acid, or vector, indicates that the cell, or nucleic acid, or vector, has been modified by the introduction of a heterologous nucleic acid or the alteration of a native nucleic acid, or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under expressed or not expressed at all. Further, the expression "recombinant" also relates to a cell, wherein further regulatory elements have been included in order to initiate or enhance expression of an otherwise silent endogenous gene, or wherein a manipulation of the regulatory elements have been performed for the same purpose. 10 (For an example of such a gene activation technique, see e.g. *Genetic Engineering News*, April 15, 1994.) 15

The term "identical" in the context of two nucleic acid or polypeptide sequences refers to the residues in the two sequences which are the same when aligned for maximum correspondence. Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith and Waterman (1981) *Adv. Appl. Math.* 2: 482, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized 20 implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI) or by inspection. 25

An additional algorithm that is suitable for determining sequence similarity is the BLAST algorithm, which is described in Altschul *et al.* (1990) *J. Mol. Biol.* 215: 30 403-410. Software for performing BLAST analyses is publicly available through the

National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>).
(See Henikoff and Henikoff (1992) *Proc. Natl. Acad. Sci. USA* 89: 10915-10919;
and Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90: 5873-5787; for
further information in this context.)

5 The term "substantial identity" or "substantial similarity" in the context of a
polypeptide indicates that a polypeptides comprises a sequence with at least 70%
sequence identity to a reference sequence, or preferably 80%, or more preferably
85% sequence identity to the reference sequence, or most preferably 90% identity
10 over a comparison window of about 10-20 amino acid residues. An indication that
two polypeptide sequences are substantially identical is that one peptide is
immunologically reactive with antibodies raised against the second peptide. Thus, a
polypeptide is substantially identical to a second polypeptide, for example, where
the two peptides differ only by a conservative substitution.

15 An indication that two nucleic acid sequences are substantially identical is that the
polypeptide which the first nucleic acid encodes is immunologically cross reactive
with the polypeptide encoded by the second nucleic acid. Another indication that
two nucleic acid sequences are substantially identical is that the two molecules
20 hybridize to each other under stringent conditions.

"Bind(s) substantially" refers to complementary hybridization between a probe
nucleic acid and a target nucleic acid and embraces minor mismatches that can be
accomodated by reducing the stringency of the hybridization media to achieve the
25 desired detection of the target polynucleotide sequence.

30 The term "antibody" refers to a polypeptide substantially encoded by an
immunoglobulin gene or immunoglobulin genes, or fragments thereof which
specifically bind and recognize an analyte (antigen). The recognized
immunoglobulin genes include the kappa, lambda, alpha, gamma, delta, epsilon and
mu constant region genes, as well as the myriad immunoglobulin variable region

genes. Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu alpha, delta, or epsilon, which in turn define the immunoglobulin classes, IgG, IgM, IgA, IgD and IgE, respectively. (In this context, see *Fundamental Immunology*, Third Edition, W.E. Paul, ed., Raven Press, N.Y. 1993).

A "chimeric antibody" is an antibody molecule in which (a) the constant region, or a portion thereof, is altered, replaced or exchanged so that the antigen binding site (variable region) is linked to a constant region of a different or altered class, effector function and/or species, or an entirely different molecule which confers new properties to the chimeric antibody, e.g., an enzyme, toxin, hormone, growth factor, drug, etc.; or (b) the variable region, or a portion thereof, is altered, replaced or exchanged with a variable region having a different or altered antigen specificity.

The term "immunoassay" is an assay that utilizes an antibody to specifically bind an analyte. The immunoassay is characterized by the use of specific binding properties of a particular antibody to isolate, target, and/or quantify the analyte.

The terms "isolated" "purified" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native state.

The phrases "specifically binds to a protein" or "specifically immunoreactive with", when referring to an antibody refers to a binding reaction which is determinative of the presence of the protein in the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind preferentially to a particular protein and do not bind in a significant amount to other proteins present in the sample. Specific binding to a protein under such conditions requires an antibody that is selected for its specificity for a particular protein. A variety of immunoassay formats may be used to select antibodies specifically immunoreactive with a particular protein. For example, solid-phase ELISA immunoassays are routinely used to select monoclonal

antibodies specifically immunoreactive with a protein. See Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbour Publications, New York, for a description of immunoassay formats and conditions that can be used to determine specific immunoreactivity.

5 A "conservative substitution", when describing a protein, refers to a change in the amino acid composition of the protein that does not substantially alter the protein's activity. Thus, "conservatively modified variations" of a particular amino acid sequence refers to amino acid substitutions of those amino acids that are not critical
10 for protein activity or substitution of amino acids with other amino acids having similar properties (e.g., acidic, basic, positively or negatively charged, polar or non-polar, etc.) such that the substitutions of even critical amino acids do not substantially alter activity. See e.g., Creighton (1984) *Protein*, W.H. Freeman and Company. In addition, individual substitutions, deletions or additions which alter,
15 add or delete a single amino acid or a small percentage of amino acids in an encoded sequence are also "conservatively modified variations".

A "gene product", as used herein, refers to a nucleic acid whose presence, absence, quantity, or nucleic acid sequence is indicative of a presence, absence, quantity, or
20 nucleic acid composition of the gene. Gene products thus include, but are not limited to, an mRNA transcript, a cDNA reverse transcribed from an mRNA, and RNA transcribed from that cDNA, a DNA amplified from the cDNA, an RNA transcribed from the amplified DNA or subsequences of any of these nucleic acids. Polypeptides expressed by the gene or subsequences thereof are also gene products.
25 The particular type of gene product will be evident from the context of the usage of the term.

Detailed description of the invention

30 In a first aspect, the present invention relates to an isolated mite-derived protein, which is an antigen derived from the mite *Sarcoptes scabiei*. Specifically, the protein according to the invention is comprised of parts or all of the sequence

disclosed in SEQ ID NO. 2. More specifically, the invention relates to a protein comprised of at least about 83 amino acids of said sequence, preferably the ones described in SEQ ID NO. 3, an analogue or a functional fragment thereof. In a specific embodiment, the protein according to the invention is comprised of at least about 100 amino acids, preferably at least about 200, such as at least about 400 amino acids of the sequence disclosed in SEQ ID NO. 2. In a specific embodiment, the protein according to the invention comprises about 400 of the last amino acids of SEQ ID NO 2, and most preferably, the sequence from about amino acid no 344 to amino acid no 770 of said sequence, which protein has been denoted Major Sarcptes Antigen 1 (MSA1) of the present inventors. In another particular embodiment, the amino acid sequence of the present protein comprises a larger part of said sequence, such as at least about 400, preferably at least about 500, e.g. at least about 600, and most preferably at least about 700, such as about 770 amino acids of said sequence, in which case it is substantially identical with the sequence disclosed in SEQ ID NO. 2. In this context, it is to be understood that all derivatives, analogues and functional fragments and functional subsequences thereof of the present proteins also fall within the scope of the invention as defined by the claims. Thus, the preferred protein according to the invention, MSA1, corresponds to a native 164 kDa protein, which provides a rapid immunogenic response in dogs and the like. Further properties of the protein according to the invention will appear from the section Experimental and the discussions below.

Thus, the above mentioned EP 0 473 111 relates to the isolation of an organism, house dust mite, which is naturally present in different surroundings than *Sarcptes scabiei*, which is a parasite in the skin of animals, such as wild foxes, swine etc., and man. More specifically, Bornstein & Hämmerling reported (Abstract World Congress of Veterinary Dermatology, 1996, p. 121) that sera from dogs that *inter alia* are oversensitive to dust mites do not present any antibodies reactive towards *Sarcptes* from red fox. Further, the clinical definition of scabies is that it is caused by *Sarcptes scabiei*, contrary to e.g. dust mite caused conditions. A further difference between *scabiei* and dust mite is that a scabies infection will give rise to

an IgG-response, contrary to dust mite allergy. Accordingly, even though there exist a certain similarity between the protein derived from the house dust mites according to EP 0 473 111 and the *Sarcoptes scabiei* protein isolated and sequenced according to the present invention, the biological roles and properties thereof differs substantially. Accordingly, there is nothing in EP 0 473 111 that would lead the skilled in this field to believe that scabies would be caused by a similar sequence. As a consequence, the pharmaceutical and diagnostic applications enabled by the EP 0 473 111 antigen are for different conditions and other subjects than the antigen according to the present invention, which mainly relates to the treatment and/or diagnosis of farm animals, such as swine, or dogs. Accordingly, even though there exist a certain similarity between the protein derived from the house dust mites according to EP 0 473 111 and the *Sarcoptes scabiei* protein isolated and sequenced according to the present invention, the differences in environment and conditions to be prevented and/or diagnosed are of such importance that 0 473 111 must be regarded as relating to a different field from the present invention.

In an especially advantageous embodiment, the protein according to the invention is a recombinant protein, which in addition to a functional part or all of the sequence disclosed in SEQ ID NO. 2 also comprises a tag, such as a conventional further amino acid sequence, which confers properties that facilitates purification, downstream analysis, such as Western blot, reversible immobilization, immunoprecipitation, immunofluorescence analysis *etc.* Said tag may e.g. be the peptide His6. In a particular embodiment, the present invention is a fusion protein, wherein the present protein or a functional subsequence thereof is fused with another protein, such as β -galactosidase, glutathione-S-transferase, protein A *etc.* In the context of fusion proteins, see e.g. Smith and Johnson (1988) *Gene* 67:31; Hopp *et al.* (1988) *Biotechnology* 6:1204; La Vallie *et al.* (1993) *Biotechnology* 11:187.

In a further embodiment, the present protein, or a functional subsequence thereof, is used to produce peptidomimetics, *i.e.* molecules that mimics the biological activity

of the peptide but no longer are completely peptidic in nature. Thus, peptidomimetics according to the invention are produced in order to provide molecules that are more advantageous than the peptides *per se* as regards e.g. size, bioavailability, duration of action, stability, storage, immunoreactivity *etc.* In this context, see e.g. Dean (1994), *BioEssays*, 683-687, Cohen and Shatzmiller (1993), *J. Mol. Graph.*, 11:166-173.

Further, the antigen according to the invention may be used in various other methods for design and/or identification of novel compounds, such as in combinatorial libraries. Such methods enable a modern drug laboratory to produce and screen millions of new chemical and/or biological compounds in a few weeks for a variety of uses. Of the large number of compounds produced, only the ones showing interesting biological activity are analyzed for further testing and experimentation. Methods for combinatorial libraries, see e.g. US patent nos. 5 753 187 and US 5 763 263, are well known to those of skill in this field and are easily arranged based on the present application. A further aspect of the present invention is the use of the antigen according to the invention in methods of high-throughput screening for the identification of a novel compound. Such methods are amenable to automated, cost-effective high throughput screening and have immediate application in a broad range of programs for development of diagnostic and/or pharmaceutically active compounds. Further, the present invention also encompasses any compound obtainable by, that is, reached through, the use of any one of the herein disclosed methods.

Thus, more specifically, the present invention also relates to a method of screening for protein or peptide analogues that mimic at least a part of the structure of the protein according to the invention, which comprises the steps of

- (a) producing a multiplicity of analogue structures and
- (b) selecting an analogue structure, wherein the three-dimensional configuration and spatial arrangement of one or more biologically active regions, preferably antigenic regions, remain substantially preserved.

In a specific embodiment, analogues mimicking a protein having the amino acid sequence as disclosed in SEQ ID NO 3 are screened for. Further, the invention also encompasses an analogue identified as disclosed above. Analogues identified by the present method are advantageously used for the same purposes as the proteins according to the invention, said various uses being disclosed elsewhere in the present disclosure.

The present proteins, functional subsequences thereof etc. may be synthesized using standard chemical peptide synthesis techniques. Where the desired subsequences are relatively short (e.g., when a particular antigenic determinant is desired), the molecule may be synthesized as a single contiguous polypeptide. Where larger molecules are desired, subsequences can be synthesized separately (in one or more units) and then fused by condensation of the amino terminus of one molecule with the carboxyl terminus of the other molecule thereby forming a peptide bond.

Solid phase synthesis in which the C-terminal amino acid of the sequence is attached to an insoluble support followed by sequential addition of the remaining amino acids in the sequence is the preferred method for the chemical synthesis of the polypeptides of this invention. Techniques for solid phase synthesis are described by Barany and Merrifield, *Solid-Phase Peptide Synthesis*; pp. 3-284 in *The Peptides: Analysis, Synthesis, Biology. Vol. 2: Special Methods in Peptide Synthesis, Part A.*, Merrifield, et al. *J. Am. Chem. Soc.*, 85: 2149-2156 (1963), and Stewart et al., *Solid Phase Peptide Synthesis, 2nd ed.* Pierce Chem. Co., Rockford, Ill. (1984).

In an alternative embodiment, the present proteins or subsequences thereof, are synthesized using recombinant DNA methodology. Generally this involves creating a DNA sequence that encodes the fusion protein, placing the DNA in an expression cassette under the control of a particular promoter, expressing the protein in a host, isolating the expressed protein and, if required, renaturing the protein.

DNA encoding the present proteins, or subsequences thereof, may be prepared by any suitable method, including, for example, cloning and restriction of appropriate sequences or direct chemical synthesis by methods such as the phosphotriester method of Narang *et al.* *Meth. Enzymol.* 68: 90-99 (1979); the phosphodiester method of Brown *et al.*, *Meth. Enzymol.* 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage *et al.*, *Tetra. Lett.*, 22: 1859-1862 (1981); and the solid support method of U.S. Patent No. 4,458,066.

Chemical synthesis produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the single strand as a template. One of skill would recognize that while chemical synthesis of DNA is limited to sequences of about 100 bases, longer sequences may be obtained by the ligation of shorter sequences. Similarly, subsequences may be cloned and the appropriate subsequences cleaved using appropriate restriction enzymes. The fragments may then be ligated to produce the desired DNA sequence.

In one embodiment, proteins of this invention may be cloned using DNA amplification methods such as polymerase chain reaction (PCR). Thus, for example, the nucleic acid sequence or subsequence is PCR amplified, using a sense primer containing one restriction site, *e.g.*, XhoI, and an antisense primer containing another restriction site, *e.g.*, BamHI. This will produce a nucleic acid encoding the desired sequence or subsequence and having terminal restriction sites. This nucleic acid can then be easily ligated into a vector containing a nucleic acid encoding the second molecule and having the appropriate corresponding restriction sites. Suitable PCR primers can be determined by one of skill in the art using the information provided in the appended sequence listing. Appropriate restriction sites can also be added to the nucleic acid encoding the protein or protein subsequence by site-directed mutagenesis. The plasmid containing the sequence or subsequence is cleaved with the appropriate restriction endonuclease and then ligated into the vector encoding the second molecule according to standard methods.

Accordingly, in a second aspect, the present invention relates to an isolated nucleic acid encoding the protein according to the present invention. More specifically, the present invention relates to a nucleic acid comprising parts or all of the base sequence disclosed in SEQ ID NO. 1, such as about 200-2000, *e.g.* at least about 400, such as at least about 1000, and preferably at least about 2300 bases thereof. In a specific embodiment of this aspect, the nucleic acid according to the invention comprises about the second half of the sequence of SEQ ID NO 1, preferably at least about 1200, *e.g.* 1284, and most preferably, the 1284 bases that encodes the protein denoted MSA1 which is discussed above. In another embodiment of this aspect of the invention, the nucleic acid is substantially identical with the sequence of SEQ ID NO 1. In a particularly advantageous embodiment, the present nucleic acid encodes a protein as disclosed by SEQ ID NO 3. In the present context, it is to be understood that the invention also encompasses any one of the above defined sequences which is a degenerate or variant thereof.

Further, this aspect of the invention also covers any nucleic acid, which hybridizes specifically under stringent conditions to a nucleic acid as disclosed above as well as any gene product obtained thereby. Such a hybridizing nucleic acid may *e.g.* be DNA, a genomic DNA sequence comprising introns as well as exons, RNA, such as mRNA *etc.*

The preparation of the present nucleic acids has already been described above. Thus, more specifically, the sequences are cloned, or amplified by *in vitro* methods, such as the polymerase chain reaction (PCR), the ligase chain reaction (LCR), the transcription-based amplification system (TAS), the self-sustained sequence replication system (SSR). A wide variety of cloning and *in vitro* amplification methodologies are known to persons of skill. Examples of these techniques and instructions sufficient to direct persons of skill through many cloning exercises are found in Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology* 152 Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.*

(1989) *Molecular Cloning - A Laboratory Manual* (2nd ed.) Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor Press, NY, (Sambrook *et al.*); *Current protocols in Molecular Biology*, F.M. Ausubel *et al.*, Current Protocols, a joint venture between Greene Publishing Associates, Inc. And John Wiley & Sons, Inc., (1994 Supplement) (Ausubel); Cashion *et al.*, U.S. patent number 5,017,478; and Carr, European Patent No. 0,246,864. Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods are found in Berger, Sambrook, and Ausubel, as well as Mullis *et al.*, (1987) U.S. Patent No. 4,683,202; *PCR Protocols A Guide to Methods and Applications* (Innis *et al.* Eds) Academic Press Inc. San Diego, CA (1990) (Innis); Arnheim & Levinson (October 1, 1990) *C&EN* 36-47; *The Journal of NIH Research* (1991) 3: 81-94 (Kwoh *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86: 1173; Guatelli *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87, 1874; Lomell *et al.* (1989) *J. Clin. Chem.*, 35: 1826; Landegren *et al.*, (1988) *Science*, 241: 1077-1080; Van Brunt (1990) *Biotechnology*, 8: 291-294; Wu and Wallace, (1989) *Gene*, 4: 560; and Barringer *et al.* (1990) *Gene*, 89: 117.

In one preferred embodiment, the nucleic acids according to the invention are isolated by routine cloning methods. The DNA sequence provided in SEQ ID NO 1, or a subsequence thereof, can be used to provide probes that specifically hybridize to the encoding gene, in a genomic DNA sample, *e.g.* in a Southern blot, or to the mRNA, in a total RNA sample (*e.g.*, in a Northern blot). Once the target nucleic acid has been identified (*e.g.*, in a Northern or Southern blot), it is isolated according to standard methods known to those of skill in the art (*see, e.g.*, Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual*, 2nd Ed. Vols. 1-3, Cold Spring Harbor Laboratory; Berger and Kimmel (1987) *Methods in enzymology*, Vol. 152: *Guide to Molecular Cloning Techniques*, San Diego: Academic press, Inc.; or Ausubel *et al.* (1987) *Current Protocols in Molecular Biology*, Greene Publishing and Wiley-Interscience, New York).

The nucleic acids according to the invention may be used in assays, such as in hybridization assays, as probes, in which case they are preferably labeled, in the

production of proteins according to the invention *etc.*, as is evident from other sections of the present specification.

5 A further aspect of the invention is an expression vector, such as a plasmid or a virus, such as a phage, which comprises a nucleic acid as describe above. Such vectors are *e.g.* useful for expressing the proteins according to the invention to provide immunogens for antibody production. Thus, vectors encoding the proteins are also useful for transforming cells *in vitro* or *in vivo* to express the present protein or a functional subsequence thereof. In addition to the present encoding
10 sequence, the vectors according to the invention also comprise suitable promoters, enhancers and further regulatory elements operably linked thereto (Queen *et al.* (1986) *Immunol. Rev.* 89:49). The vector may be a plasmid, virus, *etc.* Further, the vector may be an oligonucleotide, in which case the coding sequence may be accompanied by the required tagging sequences for use in methods such as
15 homologous recombination, as described in the literature.

Thus, an additional aspect of the present invention is a recombinant cell comprising a vector according to the invention. The culture of cells, including cell lines and cultured cells from tissue or blood samples, is well known in the art. (See *e.g.*
20 Freshney, *Culture of Animal Cells, A Manual of Basic Technique*, 3rd ed., Wiley-Liss, New York, NY (1994).) Cells expressing the present nucleic acid may be used to monitor expression levels of the protein according to the invention in a wide variety of contexts. The cells according to the invention may be prokaryotic, such as bacteria, *e.g.* *E. coli*, eukaryotic, such as yeast, mammalian, such as canine, porcine
25 or human, a protozoan *etc.*

In an additional aspect, the invention relates to a method for producing a protein according to the invention, or a functional subsequence thereof, which method comprises the steps of

30 (a) providing a DNA encoding the desired protein or polypeptide;

- (b) introducing said DNA in a suitable expression vector or expression cassette;
- (c) transfer of said vector or cassette into a suitable cell;
- (d) culturing said cell to obtain the desired product; and optionally
- (e) purification of the protein or polypeptide.

5

The transfer of the vector may be performed by well-known methods, depending on the type of cellular host, such as calcium chloride transfection, which is commonly used with prokaryotic cells, or calcium phosphate treatment, electroporation, lipofection, microinjection *etc.* Once expressed, the products may be purified according to standard procedures in the art, such as HPLC purification, fraction column chromatography, gel electrophoresis and the like (see *e.g.* Scopes, *Protein Purification*, Springer-Verlag, NY, 1982).

10

15

In a further aspect, the present invention relates to an antibody raised against a protein according to the invention, or a functional part thereof. The antibody according to the invention will preferably comprise at least about 10, more preferably at least 20, 40 or 50 and most preferably at least 100 or 200, or even 400 amino acids. In a specific embodiment, the antibody binds to a protein comprised of essentially all of the sequence disclosed in SEQ ID NO 2.

20

25

More specifically, the present invention relates to antibodies including individual, allelic, strain, or species variants, and fragments thereof, both in their naturally occurring (full-length) forms and in recombinant forms. Additionally, antibodies are raised to these polypeptides in either their native configurations or in non-native configurations. Anti-idiotypic and chimeric or bispecific antibodies can also be generated.

30

In order to produce the antibodies specifically reactive with polypeptides according to the invention, a number of immunogens are used. Recombinant or synthetic polypeptides of 8-15, preferably 10, amino acids in length, or greater, selected from amino acid sub-sequences of SEQ ID NO 2 are the preferred polypeptide

immunogen (antigen) for the production of monoclonal or polyclonal antibodies. In one class of preferred embodiments, an immunogenic peptide conjugate is also included as an immunogen. Naturally occurring polypeptides are also used, either in pure or impure form.

5 Recombinant polypeptides are expressed in eukaryotic or prokaryotic cells (as described above) and purified using standard techniques. The polypeptide, or a synthetic version thereof, is then injected into an animal capable of producing antibodies. Either monoclonal or polyclonal antibodies can be generated for
10 subsequent use in immunoassays to measure the presence and quantity of the polypeptide.

Methods of producing polyclonal antibodies are known to those of skill in the art, see, e.g., Coligan (1991) *Current Protocols in Immunology* Wiley/Greene, NY; and
15 Harlow and Lane (1989) *Antibodies: A Laboratory Manual* Cold Spring Harbor Press, NY).

Monoclonal antibodies are prepared from cells secreting the desired antibody. These antibodies are screened for binding to normal or modified polypeptides, or screened
20 for agonistic or antagonistic activity, e.g., activity mediated through a *suppressor of fused* protein. Specific monoclonal and polyclonal antibodies will usually bind with a K_D of at least about 0.1 mM, more usually at least about 50 μ M, and most preferably at least about 1 μ M or better.

25 In some instances, it is desirable to prepare monoclonal antibodies from various mammalian hosts, such as dogs, swine, rodents, primates, humans, etc. Description of techniques for preparing such monoclonal antibodies are found in, e.g. Stites *et al.* (eds) *Basic and Clinical Immunology* (4th ed.) Lange Medical Publications, Los Altos, CA, and references cited therein; Harlow and Lane, *supra*; Goding (1986)

Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York, NY; and Kohler and Milstein (1975) *Nature* 256: 495-497.

Other suitable techniques involve selection of libraries of recombinant antibodies in phage or similar vectors (see, e.g., Huse *et al.* (1989) *Science* 246: 1275-1281; and Ward, *et al.* (1989) *Nature* 341: 544-546; and Vaughan *et al.* (1996) *Nature Biotechnology*, 14: 309-314).

In a further aspect, the present invention relates to a protein as defined above, or a functional subsequence thereof, for use in a pharmaceutical preparation, preferably as a vaccine. Thus, the invention also relates to a vaccine preparation comprising a protein as defined above and a pharmaceutically acceptable carrier, such as an aqueous carrier. A variety of aqueous carriers may be used, e.g. buffered saline. The solution is sterile and generally free of any undesirable matter. It may contain further auxiliary substances as required to approximate physiological conditions, such as pH adjusting agents and buffers, toxicity adjusting agents, e.g. sodium acetate, sodium chloride, potassium chloride, sodium lactate *etc.* It may be prepared for injection or any other suitable route of administration, such as oral administration. For a brief review of drug delivery, see Langer, *Science* 249:527-1533 (1990).

More specifically, the vaccine preparation according to the invention is capable of provoking an immune response in a subject, such as an animal or human, and thereby preventing said subject from being infected with *Sarcoptes scabiei*, whereby sarcoptes mange or scabies is avoided. Thus, the vaccine composition according to the invention may advantageously be used to immunize animals, e.g. dogs or swine. As one example, dogs may be infected with scabies or sarcoptic mange from wild animals in the surroundings, such as red fox. Swine is another species that may be infected with scabies or sarcoptic mange. For example, when piglets are transported to novel locations during the breeding thereof, they may be contacted with other stock and the infection may then be spread. The vaccine composition may be a

killed or inactivated, attenuated, recombinant or subunit vaccine, as appropriate, depending on the prevailing conditions.

Thus, the invention also relates to a method of preventing mite diseases, preferably conditions caused by mites, such as *Sarcoptes scabiei*, in a subject, such as a human, or animal, such as a canine or porcine, subject, which method comprises the administration of a pharmaceutical preparation according to the invention to said subject in an effective dose. In a specific embodiment, said disease is sarcoptes mange or scabies.

The invention also relates to any medicinal use of the proteins and polypeptides according to the invention. As regards the pharmaceutical uses of the present polypeptides, it is to be understood, that in many cases, it may be more advantageous to use the peptidomimetics according to the invention than the original polypeptides. Such uses are also within the scope of the invention.

In one further aspect, the invention relates to various assays, wherein an antibody according to the invention is used to determine the presence and/or quantity of the polypeptide. The methods include analytical biochemical methods, such as electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), hyperdiffusion chromatography and the like as well as various immunological methods, such as fluid or gel precipitating reactions, immunodiffusion, immunoelectrophoresis, radioimmunoassay (RIA), enzyme-linked immunosorbent assays (ELISAs), immunofluorescence assays *etc.* As used herein, an immunoassay is an assay wherein an antibody is used to specifically bind to the analyte. In the present context, such an assay format as disclosed in EP 291 194 (Unilever) is advantageously used, or variants of said format specifically adapted to the present purpose. For a review of general immunoassays, see *Methods in Cell Biology*, Vol. 37: *Antibodies in Cell Biology*, Asai, ed., Academic Press, Inc., New York (1993); and *Basic and Clinical Immunology*, 7th ed., Stites & Terr, eds., (1991).

Competitive assay formats are preferred in the present context, wherein the amount of analyte, preferably an unknown quantity of antibodies in a subject, in a sample is measured indirectly by measuring the amount of added analyte, displaced from a capture agent by the analyte present in the sample. Most preferred are the enzyme-linked immunosorbent assay (ELISA) methods, in which an antibody typically is bound to an enzyme, such as peroxidase or phosphatase, which can produce colored reaction products from an appropriate buffer. Thus, it utilizes a tagged antigen molecule of known quantity to determine an unlabelled antigen of unknown quantity. Preferably, the protein according to the invention, or a suitable functional fragment thereof, is used coupled to a conventional tag, such as His6. This assay is e.g. useful to diagnose *Sarcoptes scabiei* infection in dogs.

Thus, in an ELISA format according to the invention, antibodies against the present polypeptide are detected and/or quantified, preferably in a biological sample. The sample may be any sample of biological tissue or fluid, such as blood. The sample is pretreated as necessary by dilution in a suitable buffer solution or concentrated, if desired. Any number of standard aqueous buffer solutions may be used, such as Tris or the like, at physiological pH. Samples are incubated with an excess of the protein according to the invention as antigen. After rinsing to remove any unbound antibody, the amount of bound antibody is quantitated by adding a solution of enzyme-conjugated antibody that binds to constant domains of antibodies in the sample. Excess conjugated antibody is rinsed away and the activity of the bound enzyme is determined by adding the substrate to the reaction and measuring the formation of products. As the products of the reactions used in ELISA procedures are colored, the amount of product formed can readily be determined by the intensity of the colour that has developed using a spectrophotometer. The activity of the bound enzyme is proportional to the amount of antigen-binding antibody in the sample; therefore, the original concentration of such antibodies can be estimated from a series of control assays employing known concentrations of specific antibodies.

More specifically, if a protein as defined by SEQ ID NO. 2 coupled to His6 is used, it has been shown by the present inventor that an antigen concentration of about 30-60 ng/ml is advantageous. It is essential that protein contamination is prevented in order to avoid an enhanced background. Further, the coating ratio should be adapted as appropriate in order to improve the background. Presumably, the aftercoating is not critical.

Consequently, the invention also relates to a kit for the diagnosis of a subject infected with a mite, preferably *Sarcoptes scabiei*. More specifically, the kit according to the invention is adapted for performing the above disclosed ELISA method. Such a kit will include one or more reagents for determining the presence or absence of antibodies in the subject raised against the polypeptides according to the invention. The antigen may be free or immobilized on a solid support, such as a test tube, a microtiter plate, a dipstick or the like. The kit may also comprise instructions for the use thereof. The kit may also contain means for the detection of labels, positive and negative controls, washing solutions *etc.* More specifically, a kit according to the present invention may advantageously comprise a conjugate, positive and negative control samples, a serum dilution buffer, a substrate, suitable washing solutions as well as an appropriate conjugate dilution buffer.

Detailed description of the drawings

Figure 1 shows the extent of the recombinant protein according to the invention in relation to the native protein.

Figure 2 illustrates the recombinant plasmid pPU17 used for the expression of the mite protein MSA1 according to the invention. Amp is the ampicillin resistance gene, lacI is the lac repressor gene, and malE is the maltose binding protein in fusion with the present MSA1.

Figure 3 is an overview of the cloning strategy of the 5' end of MSA.

Figure 4 shows the results of Western blot analysis of the mite recombinant protein MSA1 according to the invention. The protein was expressed in *E.coli*, purified and

separated by SDS-PAGE before transfer to nitrocellulose. Lanes 1, 2 and 4 show the results after analysis with positive dog sera, and lanes 3, 5 and 6 shows the results after analysis with negative dog sera. Lane M is the molecular weight markers.

5 EXPERIMENTAL

Material and methods

Collection of parasites

Living mites of both sexes and different developmental stages of *S. scabiei* were isolated from the skin of wild red foxes as described in Bornstein & Zakrisson (1993) *Vet. Dermatol.* 4:107. Briefly, pieces of skin with the fur cut down to about 10 mm were left in Petri dishes at room temperature under an electric light. Mites that migrated onto the underside of the lid were collected and stored at -70°C until further use.

Immunosera

S. scabiei proteins of a size of above 90 kD were size-selected on 10% gels by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and electroeluted as previously described, see Jacobs & Clad (1986) *Anal. Biochem.* 154:583. New Zealand White rabbits were immunized with the selected proteins and Freund's complete adjuvant.

RNA preparation and construction of cDNA library

A total of 160 mg of mites was washed with PBS and then homogenized in a glass grinder at 4°C. Total RNA was extracted with an RNAagents® kit from Promega and mRNA was isolated by oligo(dT) cellulose chromatography (Pharmacia Biotech). Double-stranded cDNA was synthesized from about 5 µg *S.scabiei* mRNA (ZAP-cDNA, Stratagene), portions ligated into UNI-ZAP XR vector (Stratagene) and packaged into λ-phage (Gigapack Gold II, Stratagene).

Cloning and sequencing of MSA1

The amplified library was screened with sera from the rabbits previously immunized with gel purified antigens. Immuno reactive plaques were cloned and the phages eluted. From a positive λ -phage clone, a pBluescript SK-subclone was generated using the ExAssist™ phage system (Stratagene). The resulting clone was designated pPU3 and the *S. scabiei* cDNA insert was mapped for positions of restriction endonuclease cleavage using enzymes from Pharmacia Biotech (Uppsala, Sweden) and New England Biolabs. For the sequencing of pPU3, nested deletions and cDNA sequence specific primers were used in conjunction with the T7 sequence kit from Pharmacia Biotech.

Subcloning and expression of MSA1

Construction of high expression vector

The open reading frame of the recombinant plasmid pPU3 was amplified by PCR using *Pfu* DNA polymerase (Stratagene) and subcloned into the XmnI-XbaI sites of the vector pPU16. The pPU16 vector is a derivative of the pMal-c2 vector (New England Biolabs) that has a sequence of DNA encoding for 6 consecutive histidine residues and a stop codon between the PstI and HindIII site. The resulting plasmid pPU17 encodes for the recombinant mite protein with the maltose binding protein fused to its N-terminus and a His6-tag fused to its C-terminus. The junction between the maltose binding protein and the recombinant mite protein has a factor Xa-binding site (IEGR).

Expression as fused protein

The recombinant plasmid pPU17 was transformed into *E. coli* strain BL21 (DE3). The resulting transformant was inoculated into 10 ml minimal medium with casamino acids and heavy metals (MM/CA) (Pryor & Leiting, Protein Expr Purif 10:309) containing ampicillin (100 mg/ml) and grown overnight at 37°C with

continuous shaking. Five ml of the overnight culture was diluted in 500 ml fresh MM/CA medium with ampicillin (100 mg/ml) and grown in a 2-liter Ehrlenmeyer flask shaking efficiently at 37°C until the OD value at 600 nm became 0.8. The culture was then cooled to 18°C, induced with 0.5 mM IPTG and then transferred to a shaker at 18°C for continued growth for 20 h. After the expression the cells were collected by centrifugation at 4000xg for 20 min and resuspended in 25 ml 20 mM phosphate buffer (pH 7.6) with 0.5 M NaCl and 30 mM imidazole, supplemented with the Complete™ protease inhibitor (Roche) and frozen overnight at -70°C. The frozen cell suspension was thawed in cold water, placed in an ice-water bath and sonicated. The suspension was clarified by centrifugation at 9000xg for 30 min and the resulting supernatant was used for affinity purification.

Minimal medium with casamino acids and heavy metal (MM/CA) is: 5 g/l glucose, 1 mg/l (+) biotin, 2 mg/l thiamine, 1 g/l (NH₄)₂SO₄, 4 ml of a 250xheavy metal stock solution, 50 ml sterile 10% casamino acids, and 200 ml sterile 5xphosphate buffer. One liter of 250xheavy metal stock solution is 500 mg MoNa₂O₄2H₂O, 250 mg CoCl₂, 175 mg CuSO₄5 H₂O, 1 g MnSO₄ H₂O, 8.75 g MgSO₄7 H₂O, 1.25 g ZnSO₄7 H₂O, 1.25 g FeCl₂4 H₂O, 2.5 g CaCl₂2 H₂O, and 1.0 g H₃BO₃ in 1 M HCl. One liter of 5xphosphate buffer contains 53 g K₂HPO₄ and 24.7 g KH₂PO₄.

Purification of recombinant mite antigen fusion protein

The supernatant was loaded onto a 1 ml HiTrap® chelating column (Amersham Pharmacia Biotech) loaded with N²⁺ at a flow rate of 1 ml/min. After washing with 10 ml of 20 mM phosphate buffer (pH 7.6) with 0.5 M NaCl and 30 mM imidazole, the captured recombinant protein was eluted with 3 ml 20 mM phosphate buffer (pH 7.6) with 0.5 M NaCl and 500 mM imidazole. After buffer exchange to 20mM Tris-HCl (pH 8.0) with 150 mM NaCl and 1mM EDTA, the maltose binding protein was cleaved off with factor Xa at room temperature for 20 h after which the recombinant mite protein was purified on a HiTrap® Q column. As preserver

Micro-O-Protect (Roche) was added to the purified recombinant protein in concentration of 0.1 %.

Cloning of part of the 5' cDNA end of MSA1

5 A PCR strategy was used in order to clone regions upstream of the cDNA insert in pPU3. In the first PCR the primer KBE 5 (5'CAC TAT CGG AGA ACG TAA CTT CGG 3'), complementary to the anti-sense strand of the insert in pPU3, was designed and used together with a T3 primer, complementary to the vector used to construct the cDNA library. As a template the *S. scabiei* cDNA-library was used.

10 The resulting fragment was cloned into the *Sma*I-site of pUC18 and sequenced as above. This new fragment was then used for the design of a second primer KBE 8 (5'CCT GGC ATT CTA CTT GAG ATG TA 3') for the amplification an additional 5'end cDNA fragment. The second 5'end fragment was cloned and sequenced as above. A continuous cDNA which included the original MSA1 cDNA and both of the

15 5' end fragments was generated by using the Titan™ One Tube RT-PCR system (manufactured by Roche). For the reverse transcriptase step the reverse primer MSA1Xba (5'CGC TCT AGA CTC AAC AAT GAA TGT CTG CAA 3') was used. In the PCR, the reverse primer was used in combination with the forward primer LDL 2 (5'CGG GAT CCG AAT ATT TCG TCT CGA AAC CG 3'). The resulting

20 fragment was cloned into the *Bam*HI-*Xba*I sites of pPU16 utilizing the recognition sites introduced during the PCR (shown in boldface). A graphic overview of the cloning strategy is shown in Fig.

Enzyme-linked immunosorbent assay

25 The recombinant mite protein MSA1 was diluted in a 0.1 M carbonate buffer, pH 9.6, to a concentration of 62.5 ng/ml and coated overnight at 4°C onto microtitre plates (Polysorp, manufactured by Nunc) in volumes of 100 µl per well. The plates was washed once with phosphate buffered saline with 0.05% Tween20 (PBS-T) followed by the addition of serum samples at a 1:100 dilution in PBS-T. After a 1h

30 incubation at 37°C the plates were washed three times with PBS-T and a monoclonal

mouse anti-dog IgG antibody, dilution 1:1000, and rabbit anti-mouse Ig conjugated to HRP (manufactured by Dako), dilution 1:1000, in PBS-T plus 1% normal rabbit serum were added to the wells and incubated for 1h at 37°C. After washing three times with PBS-T, 100µl of TMB (manufactured by Sigma) were added to each well. The substrate incubation was stopped with 50 µl of H₂SO₄ and the amount of end product was analyzed at 450 nm in Dynatech MR5000 spectrophotometer (manufactured by Dynatech).

Affinity purification of antibodies

Purified recombinant mite protein or recombinant phages in *E. coli* expressing the recombinant mite protein were transferred to a nitro cellulose filter. After blocking the filter with 1% BSA (Stratagene) in 20 mM Tris-HCl, 0.5 M NaCl, 0.05% Tween20, pH 7.5 (TTBS), anti-*S. scabiei* rabbit serum was added to the filter and incubated for 1h at room temperature. After extensive washing with TTBS, the bound antibodies were eluted and used in Western blot analysis on *S. scabiei* antigens (Beall & Mitchell, J. Immunol Method. (1986) 86:217). The affinity purified antibodies were also used to localize the protein in *S. scabiei* mites.

Western blot analysis

Purified recombinant mite proteins were separated by SDS-PAGE in 12% mini gels in Mini PROTEAN II cell (Bio-Rad Laboratories). Blotting of proteins to nitrocellulose was done by using an electrophoretic transfer cell (Bio-Rad Laboratories). After completion of transfer, the membrane bound proteins were checked by briefly incubating the membrane in a 0.2% Ponceau-S solution (Sigma) followed by rinsing in distilled water. The blots were blocked with phosphate buffer saline (PBS) containing non-fat dry milk, incubated with sera from experimentally dogs (Bornstein & Zakrisson (1993) *Vet. Dermatol.* 4:107), then incubated with a mouse anti-dog IgG monoclonal antibody and a rabbit anti-mouse IgG. After washing, bound antibodies were visualized by chemiluminescence detection and exposure to film using the ECL-system (Amersham Pharmacia Biotech).

CLAIMS

1. An isolated mite protein comprising of at least about 83 amino acids of the sequence disclosed in SEQ ID NO. 2, said 83 amino acids being essentially identical to the amino acid sequence disclosed in SEQ ID NO 3.
2. A protein according to claim 1, which comprises at least about 400, such as about 427, amino acids of the sequence disclosed in SEQ ID NO. 2.
3. A protein according to claim 2, which is comprised of the last about 427 amino acids of the sequence disclosed in SEQ ID NO. 2.
4. An isolated nucleic acid encoding a protein according to any one of claims 1-3.
5. A nucleic acid according to claim 4, the nucleotide sequence of which is substantially identical with bases no 1030-1279 of the sequence disclosed in SEQ ID NO. 1.
6. A nucleic acid which hybridizes specifically under stringent conditions to a nucleic acid according to claim 4 or 5.
7. An expression vector which comprises a nucleic acid according to any one of claims 4-6.
8. A recombinant cell comprising a vector according to claim 7.
9. A method for producing a protein, which method comprises the steps of
 - (a) providing a DNA according to any one of claims 4-6;
 - (b) introducing said DNA in an expression vector;
 - (c) insertion of said vector into a suitable host cell;
 - (d) culturing said host cell to obtain the desired protein product; and optionally
 - (e) purification of the protein or polypeptide produced.
10. An antibody raised against a protein according to any one of claims 1-3.
11. An antibody according to claim 10, which is a monoclonal antibody.
12. Use of a protein according to any one of claims 1-3 in an immunosorbent assay, such as enzyme-linked immunosorbent assay (ELISA).
13. Use of a protein according to any one of claims 1-3 in a screening method wherein compounds having the same or similar biological activities as said protein are identified.

14. A method for screening protein or peptide analogues that mimic at least a part of the structure of the protein according to any one of claims 1-3, which comprises the steps of
- (a) producing a multiplicity of analogue structures and
- 5 (b) selecting an analogue structure, wherein the three-dimensional configuration and spatial arrangement of one or more biologically active regions remain substantially preserved.
15. A method according to claim 14, wherein analogues mimicking a protein having the amino acid sequence essentially as disclosed in SEQ ID NO 3 are screened for.
- 10 16. A protein according to any one of claims 1-3 for use as a vaccine.
17. Use of a protein according to any one of claims 1-3 in the manufacture of a vaccine preparation.
18. A vaccine preparation comprising a protein according to any one of claims 1-3 and a pharmaceutically and/or veterinary acceptable carrier.
- 15 19. A vaccine preparation according to claim 18 for the prevention of *Sarcoptes mangle* or scabies.
20. A method of preventing a disease associated with mites, such as *Sarcoptes scabiei*, in a subject, such as a human, canine or porcine subject, which method comprises administration of a preparation according to claim 18 or 19 to said subject in a pharmaceutically effective dose.
- 20 21. A method according to claim 20, wherein said disease is *sarcoptes mangle* or scabies.
22. A method for the diagnosis of a mite associated disease comprising the steps of
- 25 a) immobilising a protein according to any one of claims 1-3;
- b) providing a sample suspected of being infected with said mite associated disease;
- c) incubation of said sample with said immobilised protein; and
- d) detection of any antibody bound to the immobilised antigen and thus specific for said mite associated disease; whereby a conclusion regarding the diagnosed condition is obtained.
- 30

23. A method according to claim 22, wherein the mite associated disease is sarcoptic mange or scabies.

24. A kit for performing the method according to claim 22 or 23.

WO 00/50450

PCT/SE00/00346

1/4

Fig. 1

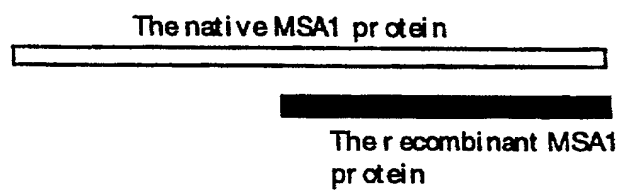
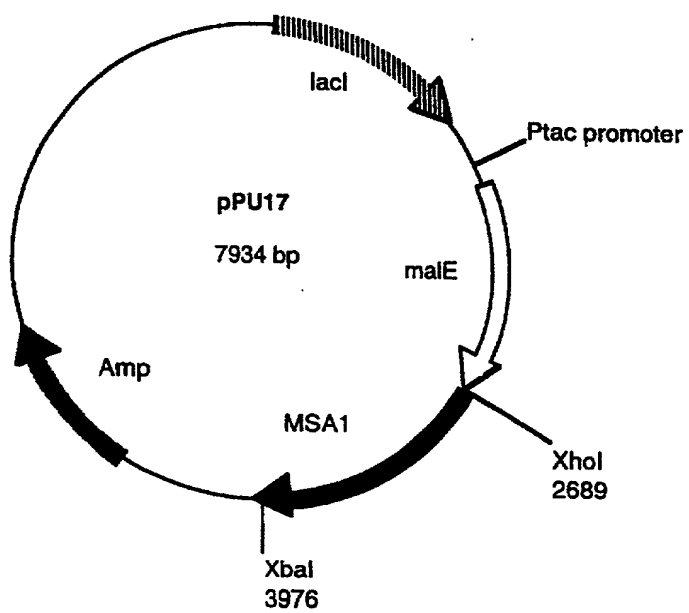
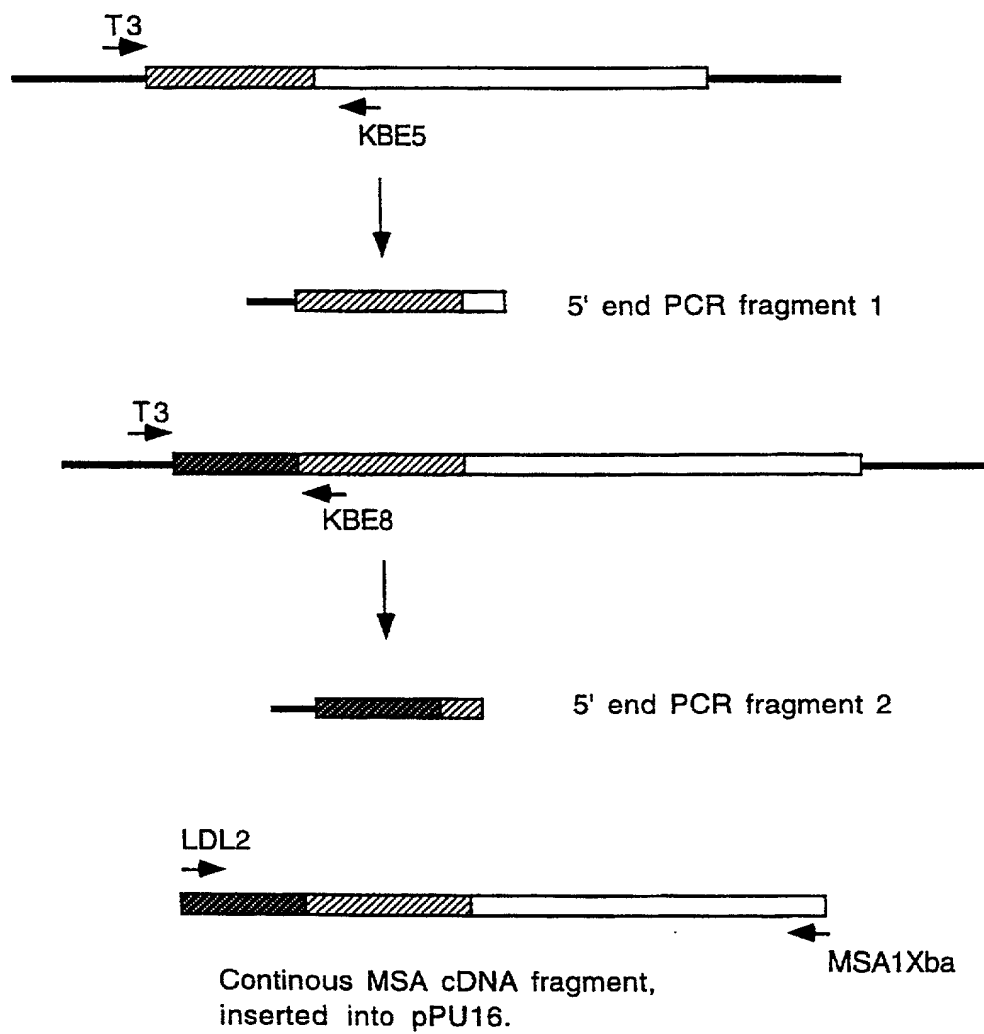


Fig. 2



3/4

Fig. 3

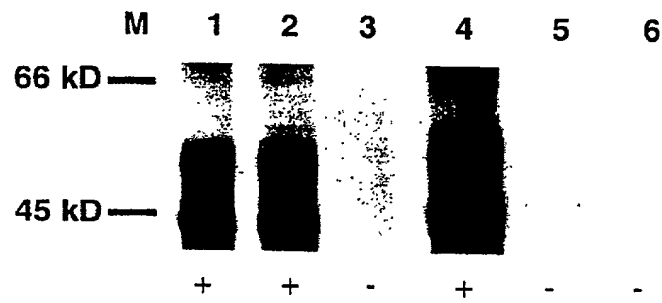


WO 00/50450

PCT/SE00/00346

4/4

Fig. 4



COMBINED DECLARATION AND POWER OF ATTORNEY #6

As a below named inventor, I hereby declare that

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

MITE PROTEIN

23

the specification of which: *(check one)*

REGULAR OR DESIGN APPLICATION

☐ is attached hereto.

☐ was filed on _____ as application Serial No. _____ and was amended on (if applicable).

PCT FILED APPLICATION ENTERING NATIONAL STAGE

☒ was described and claimed in International application PCT/SE00/00346 filed on 22 February 2000 and as amended on (if any).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

PRIORITY CLAIM

I hereby claim foreign priority benefits under 35 USC 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

PRIOR FOREIGN APPLICATION(S)

| Country | Application Number | Date of Filing (day, month, year) | Priority Claimed |
|---------|--------------------|--------------------------------------|------------------|
| Sweden | 9900674-4 | 25 February 1999 | yes |
| | | | |

(Complete this part only if this is a continuing application.)

I hereby claim the benefit under 35 USC 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37 Code of Federal Regulations §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

(Application Serial No.)

(Filing Date)

(Status--patented, pending, abandoned)

POWER OF ATTORNEY

The undersigned hereby authorizes the U.S. attorney or agent named herein to accept and follow instructions from Albihns Stockholm AB as to any action to be taken in the Patent and Trademark Office regarding this application without direct communication between the U.S. attorney or agent and the undersigned. In the event of a change in the persons from whom instructions may be taken, the U.S. attorney or agent named herein will be so notified by the undersigned.

As a named inventor, I hereby appoint the registered patent attorneys represented by Customer No. 000466 to prosecute this application and transact all business in the Patent and Trademark Office connected therewith, including: Robert J. PATCH, Reg. No. 17,355, Andrew J. PATCH, Reg. No. 32,925, Robert F. HARGEST, Reg. No. 25,590, Benoit CASTEL, Reg. No. 35,041, Eric JENSEN, Reg. No. 37,855, Thomas W. PERKINS, Reg. No. 33,027, and Roland E. LONG, Jr., Reg. No. 41,949,

c/o YOUNG & THOMPSON,
Second Floor,
745 South 23rd Street,
Arlington, Virginia 22202.

00466

PATENT TRADEMARK OFFICE

Address all telephone calls to Young & Thompson at 703/521-2297. Telefax: 703/685-0573.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor: Jens MATTSSON
(given name, family name)

Inventor's signature

Date

27/8 2001Residence: Uppsala, Sweden SECitizenship: Swedish

Post Office Address: Odlingsvagen 2A
SE-756 55 Uppsala, Sweden

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: NATIONAL VETERINARY INSTITUTE
- (B) STREET: P.O. Box 7073
- (C) CITY: UPPSALA
- (E) COUNTRY: SWEDEN
- (F) POSTAL CODE (ZIP): 750 07
- (G) TELEPHONE: +46 18 67 40 00
- (H) TELEFAX: +46 18 30 91 62

(ii) TITLE OF INVENTION: MITE PROTEIN

(iii) NUMBER OF SEQUENCES: 3

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1284 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: YES

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1284

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

| | |
|---|-----|
| GAA GCG GAA GTT ACG TTC TCC GAT AGT GAA GAT AAG AAA AAT TAT TTC | 48 |
| Glu Ala Glu Val Thr Phe Ser Asp Ser Glu Asp Lys Lys Asn Tyr Phe | |
| 1 5 10 15 | |
| GTC GAA CTT AAA AAA GAT AAA GAT TTA TAT TCG ATG AAA TCG AAT GTG | 96 |
| Val Glu Leu Lys Lys Asp Lys Asp Leu Tyr Ser Met Lys Ser Asn Val | |
| 20 25 30 | |
| AAA CGA AAC AAT GAG ATT TTC TAT GAG AAC AAT ATG GAT TTG GAG AAG | 144 |
| Lys Arg Asn Asn Glu Ile Phe Tyr Glu Asn Asn Met Asp Leu Glu Lys | |
| 35 40 45 | |

Patent 2544660

| | |
|---|-----|
| AAC GGT AAA ATG AAT TGG TAT TAC AAA CGA AAC GAT CGA ACA TGG AAT Asn Gly Lys Met Asn Trp Tyr Tyr Lys Arg Asn Asp Arg Thr Trp Asn 50 55 60 | 192 |
| ATG GAT CTC GAT AAT GCA TTC AAT CCA AGA GAT GGT ACA ATG AAA CTT Met Asp Leu Asp Asn Ala Phe Asn Pro Arg Asp Gly Thr Met Lys Leu 65 70 75 80 | 240 |
| CAA GTG AAA GAT CGT ATC TAT GAT ATC AAA TTG AAA CGA GAA CCG TTC Gln Val Lys Asp Arg Ile Tyr Asp Ile Lys Leu Lys Arg Glu Pro Phe 85 90 95 | 288 |
| CGA TAC GGT GAT CTA CAT ATC GAA GGA AAT GAG AAT CCT TTG ATC AAA Arg Tyr Gly Asp Leu His Ile Glu Gly Asn Glu Asn Pro Leu Ile Lys 100 105 110 | 336 |
| AAG GGT GAT TTA CAT ATG TCT CTC GTC GAT CCG CTT ACT TTG AAT GTT Lys Gly Asp Leu His Met Ser Leu Val Asp Pro Leu Thr Leu Asn Val 115 120 125 | 384 |
| TTG ACC AAG AAT GAT GGA ATC GTC GAT ATG ACA TTG GAT TTG GTC TCT Leu Thr Lys Asn Asp Gly Ile Val Asp Met Thr Leu Asp Leu Val Ser 130 135 140 | 432 |
| CCC AAC ACC AAA AAA GCA GCG CTA AAA ATC AAT TCG AAA AAA TAC GAT Pro Asn Thr Lys Lys Ala Ala Leu Lys Ile Asn Ser Lys Lys Tyr Asp 145 150 155 160 | 480 |
| CTT GAT CAT GAT GGT GAG ATT ACC GTT TCG ATC TTT AAT CCT CGA ATG Leu Asp His Asp Gly Glu Ile Thr Val Ser Ile Phe Asn Pro Arg Met 165 170 175 | 528 |
| ACT TGG AAA CAT CAC ACT AGA AAA GGT GAT ATG GAA TTG AAT ATT GAT Thr Trp Lys His His Thr Arg Lys Gly Asp Met Glu Leu Asn Ile Asp 180 185 190 | 576 |
| GCT GAT ATC ACT CGA AAA GGT TCA TTG ATC ACC TAT TT CGT AAA GAG Ala Asp Ile Thr Arg Lys Gly Ser Leu Ile Thr Tyr Ser Arg Lys Glu 195 200 205 | 624 |
| CCA GAT GAT TCG ACA AAA GTT CGA TAT TCA AGA CAA GGA AT CAA GTT Pro Asp Asp Ser Thr Lys Val Arg Tyr Ser Arg Gln Gly Asn Gln Val 210 215 220 | 672 |
| TCG ATG GAA GTC GAT TCT AAA TTG ATC GAA GGC CAT GCG AAC GAA ACT Ser Met Glu Val Asp Ser Lys Leu Ile Glu Gly His Ala Asn Gly Thr 225 230 235 240 | 720 |
| TTG ACC GAT GGC AAA ATT CAT GTC AAA GGT CGA GAG AGT GAT TTC AA Leu Thr Asp Gly Lys Ile His Val Lys Gly Arg Glu Ser Asp Phe Gly 245 250 255 | 768 |
| ATC GAA AGC ACC TAT AAA GTT GAA GAT GGT AAG CTT ATG ATT GAG CCA Ile Glu Ser Thr Tyr Lys Val Glu Asp Gly Lys Leu Met Ile Glu Pro 260 265 270 | 816 |
| ACC AAA ACT CAG AAT GGA AAA TTA GAA GGT CTT CTT TCG AGA AAA GTA Thr Lys Thr Gln Asn Gly Lys Leu Glu Gly Leu Leu Ser Arg Lys Val 275 280 285 | 864 |

000443501

CCA TCA CAT CTT GTT CTT GAA ACA CCA AGA GTG AAA ATG AAC ATG AAA 912
 Pro Ser His Leu Val Leu Glu Thr Pro Arg Val Lys Met Asn Met Lys
 290 295 300

TAT GAT AGA TTT GCT CCG GTG AAG ATA TTG AAA TTA GAT TAC GAT GGT 960
 Tyr Asp Arg Phe Ala Pro Val Lys Ile Leu Lys Leu Asp Tyr Asp Gly
 305 310 315 320

TTG AAT TAT GAG AAA CAT ATC GAT GCT GAA TAC GAG CCA TCA AAT CAT 1008
 Leu Asn Tyr Glu Lys His Ile Asp Ala Glu Tyr Glu Pro Ser Asn His
 325 330 335

TAC AAA TAC TTT ACC GAT GGT AAA TCG AAG AGA AGT GGC AAA GGT TAT 1056
 Tyr Lys Tyr Phe Thr Asp Gly Lys Ser Lys Arg Ser Gly Lys Gly Tyr
 340 345 350

TCG ATC AAA ATC GAT GGA AAA CCA AAG AAA GCA TTG AAA GTT GAT GTC 1104
 Ser Ile Lys Ile Asp Gly Lys Pro Lys Lys Ala Leu Lys Val Asp Val
 355 360 365

GAT ATG CCG GAT TTT AAA TTC AAT GTG AAC AAA CCG GAA GAT AGT AAC 1152
 Asp Met Pro Asp Phe Lys Phe Asn Val Asn Lys Pro Glu Asp Ser Asn
 370 375 380

AAA GCT CAA TTT AGT TAT ACA TTC AAT GAT TAT ACC GAA ACG GAA GAG 1200
 Lys Ala Gln Phe Ser Tyr Thr Phe Asn Asp Tyr Thr Glu Thr Glu Glu
 385 390 395 400

TAT GAA TTC GAT CCA CAT CGT GCA TAT ATC TTG AAT TGG GCC AGA GCT 1248
 Tyr Glu Phe Asp Pro His Arg Ala Tyr Ile Leu Asn Trp Ala Arg Ala
 405 410 415

ATC AGA CAA TAT TTG CAG ACA TTC ATT GTT GAG TAG 1284
 Ile Arg Gln Tyr Leu Gln Thr Phe Ile Val Glu *
 420 425

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 428 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Glu Ala Glu Val Thr Phe Ser Asp Ser Glu Asp Lys Lys Asn Tyr Phe
 1 5 10 15

Val Glu Leu Lys Lys Asp Lys Asp Leu Tyr Ser Met Lys Ser Asn Val
 20 25 30

Lys Arg Asn Asn Glu Ile Phe Tyr Glu Asn Asn Met Asp Leu Glu Lys
 35 40 45

Asn Gly Lys Met Asn Trp Tyr Tyr Lys Arg Asn Asp Arg Thr Trp Asn
 50 55 60

Met Asp Leu Asp Asn Ala Phe Asn Pro Arg Asp Gly Thr Met Lys Leu
 65 70 75 80

Gln Val Lys Asp Arg Ile Tyr Asp Ile Lys Leu Lys Arg Glu Pro Phe
 85 90 95
 Arg Tyr Gly Asp Leu His Ile Glu Gly Asn Glu Asn Pro Leu Ile Lys
 100 105 110
 Lys Gly Asp Leu His Met Ser Leu Val Asp Pro Leu Thr Leu Asn Val
 115 120 125
 Leu Thr Lys Asn Asp Gly Ile Val Asp Met Thr Leu Asp Leu Val Ser
 130 135 140
 Pro Asn Thr Lys Lys Ala Ala Leu Lys Ile Asn Ser Lys Lys Tyr Asp
 145 150 155 160
 Leu Asp His Asp Gly Glu Ile Thr Val Ser Ile Phe Asn Pro Arg Met
 165 170 175
 Thr Trp Lys His His Thr Arg Lys Gly Asp Met Glu Leu Asn Ile Asp
 180 185 190
 Ala Asp Ile Thr Arg Lys Gly Ser Leu Ile Thr Tyr Ser Arg Lys Glu
 195 200 205
 Pro Asp Asp Ser Thr Lys Val Arg Tyr Ser Arg Gln Gly Asn Gln Val
 210 215 220
 Ser Met Glu Val Asp Ser Lys Leu Ile Glu Gly His Ala Asn Gly Thr
 225 230 235 240
 Leu Thr Asp Gly Lys Ile His Val Lys Gly Arg Glu Ser Asp Phe Glu
 245 250 255
 Ile Glu Ser Thr Tyr Lys Val Glu Asp Gly Lys Leu Met Ile Glu Pro
 260 265 270
 Thr Lys Thr Gln Asn Gly Lys Leu Glu Gly Leu Leu Ser Arg Lys Val
 275 280 285
 Pro Ser His Leu Val Leu Glu Thr Pro Arg Val Lys Met Asn Met Lys
 290 295 300
 Tyr Asp Arg Phe Ala Pro Val Lys Ile Leu Lys Leu Asp Tyr Asp Gly
 305 310 315 320
 Leu Asn Tyr Glu Lys His Ile Asp Ala Glu Tyr Glu Pro Ser Asn His
 325 330 335
 Tyr Lys Tyr Phe Thr Asp Gly Lys Ser Lys Arg Ser Gly Lys Gly Tyr
 340 345 350
 Ser Ile Lys Ile Asp Gly Lys Pro Lys Lys Ala Leu Lys Val Asp Val
 355 360 365
 Asp Met Pro Asp Phe Lys Phe Asn Val Asn Lys Pro Glu Asp Ser Asn
 370 375 380
 Lys Ala Gln Phe Ser Tyr Thr Phe Asn Asp Tyr Thr Glu Thr Glu Glu
 385 390 395 400

Tyr Glu Phe Asp Pro His Arg Ala Tyr Ile Leu Asn Trp Ala Arg Ala
 405 410 415

Ile Arg Gln Tyr Leu Gln Thr Phe Ile Val Glu *
 420 425

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: YES

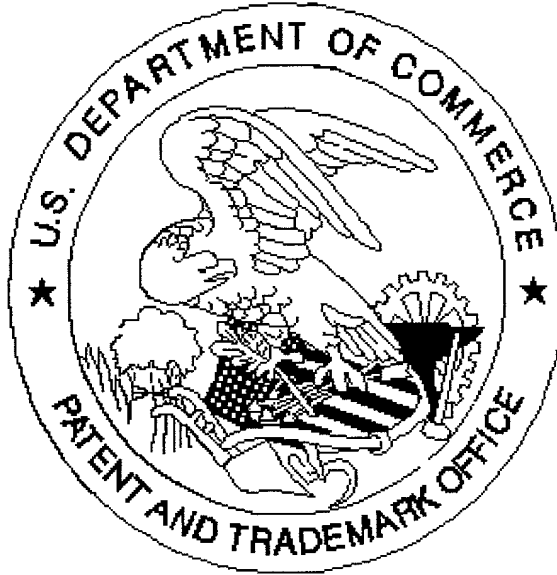
(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

Ser Arg Cys Asp Leu Gln His His His His His His His
 1 5 10

FOI 254760

United States Patent & Trademark Office
Office of Initial Patent Examination -- Scanning Division



Application deficiencies found during scanning:

☐ Page(s) _____ of _____ were not present
for scanning. (Document title)

☐ Page(s) _____ of _____ were not
present
for scanning. (Document title)

☒ Scanned copy is best available. Lines are crossing
all pages of this application.